

Advances in Clotting Factors: From Bench to Bedside

Anthony Sung, MD

Duke University Medical Center, Durham, North Carolina

Abstract The history of clotting factors is inextricably tied to that of hemophilia. The development of recombinant factor VIII (rFVIII) and factor IX (rFIX) in the 1990s has resulted in hemophilia patients having a virtually normal lifespan and significantly fewer complications, such as joint and intracranial bleeding, following prophylactic infusions. However, these treatments are limited by the short-half lives of rFVIII and rFIX, meaning that patients must endure three- or four-times-weekly infusions. This report summarizes some of the exciting advances in the management of hemophilia that have been reported in recent years. They include the addition of polyethylene glycol (PEGylation) or fusion with another protein, such as immunoglobulin G or albumin, to decrease the frequency of infusions, provide prolonged protection from bleeding, limit the need for central venous access devices, and encourage patients to transition to a prophylaxis regimen. In addition, advances have been made in improving the safety and efficacy of long-lasting rFVIII proteins for hemophilia A and the development of alternative agents to treat hemophilia and anticoagulant-related bleeding (eg, warfarin reversal) and new oral anticoagulants (eg, direct thrombin or factor Xa inhibitors).

Hemophilia is caused by functional deficiency of a single coagulation protein. Absence of such proteins—factor VIII (FVIII) in hemophilia A and factor IX (FIX) in hemophilia B—may lead to spontaneous internal bleeding with joint damage, intracranial hemorrhage, and death. Since the 1840s, transfusion of whole blood has been used to treat hemophilia-associated bleeding.¹ It wasn't until 1911 that FVIII was detected in plasma² and 1937 when its role was described in hemostasis and the coagulation cascade³ (Figure 1). These advances led to the development of plasma transfusions in the 1940s, plasma concentrates in the 1950s, cryoprecipitate in the early 1960s, and freeze-dried FVIII products for storage and home use in the late 1960s.⁴

However, as use of blood products became more common, concerns began to arise about infectious contamination, highlighted by outbreaks of hepatitis C

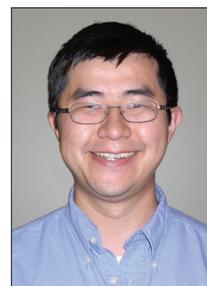
virus infection in the 1970s and human immunodeficiency virus (HIV) infection in the 1980s among hemophilia patients receiving pooled blood products. Concerns about infected blood products led to the development of recombinant FVIII (rFVIII; approved by the US Food and Drug Administration in 1992) and recombinant FIX (rFIX; approved in 1997).⁴

The use of these recombinant coagulation proteins has changed hemophilia care and increased the life-expectancy of patients with severe hemophilia from approximately 20 years in the 1970s to an essentially normal lifespan today. Long-term prophylactic factor replacement therapy also has reduced morbidity, decreasing the risk of joint damage and intracranial hemorrhage, and improved the quality of life for both children and adults with hemophilia.⁵⁻⁷ Recombinant factors are safe and provide independence from reliance on donor blood products.

However, use of recombinant factors

is not without its challenges. These substances are expensive, costing more than \$250,000 a year per adult in the United States.⁴ Because of their short half-life, they need to be administered every other day or several times a week, representing a significant time commitment and presenting problems with maintaining adherence. In addition, they require venous access, which often requires placement of a central venous access device (ie, port) and involves risks of sepsis and thrombosis. Finally, 25%–30% of hemophilia A and 3%–4% of hemophilia B patients develop alloantibodies to these recombinant factors, inhibiting their efficacy and resulting in renewed problems with bleeding.⁸ Whereas gene therapy is a potential solution,⁹ a cure is not yet available.

Thus, there is an urgent need for new, improved clotting factors to treat hemophilia patients and individuals with acquired bleeding problems due to administration of oral anticoagulants (eg, warfarin) who either have been given a supratherapeutic dose or have developed bleeding in conjunction with trauma and surgery. New oral anticoagulants, such as rivaroxaban and dabigatran, also present challenges to reversal and inhibition of bleeding events. Prothrombin complex concentrates (PCCs), fresh frozen plasma (FFP), anti-inhibitor coagulant complex



Dr. Sung is a Fellow in Hematology/Oncology at Duke University Medical Center, Durham, North Carolina.

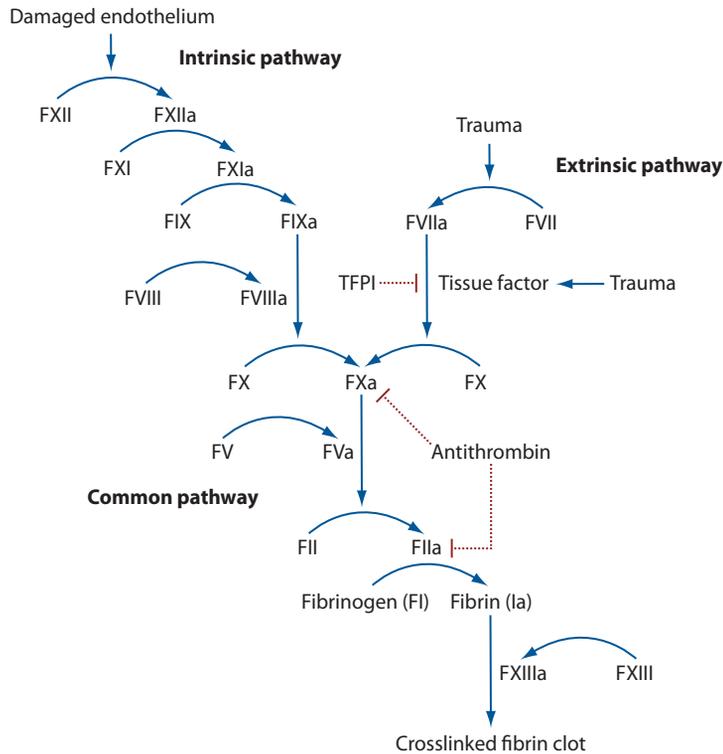


FIGURE 1 Schematic representation of the coagulation cascade, showing the role and importance of the various clotting factors as well as the site of action of tissue factor pathway inhibitor (TFPI) and antithrombin. FXII = factor XII (Hageman factor); FXIIa = activated FXII; FXI = factor XI (deficient in hemophilia C); FXIa = activated FXI; FIX = factor IX (Christmas factor; deficient in hemophilia B); FIXa = activated FIX; FVIII = factor VIII (Brinkhous factor, deficient in hemophilia A); FVIIIa = activated FVIII; FX = factor X (Stuart-Prower factor); FXa = activated FX; FVII = factor VII (stable factor); FVIIa = activated FVII; FV = factor V (proaccelerin); FVa = activated FV; FII = factor II (prothrombin); FIIa = activated factor II (thrombin); FXIII = factor XIII (Laki-Lorand factor); FXIIIa = activated FXIII.

(factor eight inhibitor bypassing activity; FEIBA), and recombinant activated factor VII (rFVIIa) are being tested in the setting of traumatic or surgical bleeding and to reverse bleeding complications resulting from the use of oral anticoagulants.

MOLECULAR APPROACHES FOR IMPROVED CLOTTING FACTORS FOR HEMOPHILIA

Based on a presentation by Randal J. Kaufman, PhD, Professor, Del E. Webb Center for Neuroscience, Aging, and Stem Cell Research, and Director, Degenerative Disease Research Program, Sanford-Burnham Medical Research Institute, La Jolla, California.

Although the development of recombinant factors in the 1990s represented a tremendous advance in the treatment of hemophilia, efforts have been underway to circumvent problems associated with

their short half-life and frequent dosing. Research teams have reported on the structure of FVIII and FIX and their mechanisms of action, allowing engineering of variants with improved half-lives. These methods include conjugation with polyethylene glycol (PEG) in a process known as PEGylation, use of PEGylated liposomes as a mechanism for sustained release, fusion with the crystallizable fragment (Fc) of the constant region of immunoglobulin (Ig) or albumin, and novel modifications to clotting factors. Although PEGylated liposomes have failed to demonstrate *in vivo* efficacy,¹⁰⁻¹² other methods have shown considerable promise.

PEGylation

PEGylation is achieved by covalently attaching PEG to residues on target proteins such as lysine or N-terminal amines.

However, random PEGylation may reduce the activity of the protein, and product heterogeneity may result in inconsistent effectiveness.

More recently, site-directed PEGylation via attachment of PEG-maleimide to cysteine residues has improved results with many proteins, including tumor necrosis factor- α , monoclonal antibody Fab fragment, vascular endothelial growth factor-aptamer, epoetin β , and interferon α .^{13,14} In this approach, missense mutations are introduced at surface residues of FVIII to incorporate cysteine residues for conjugation with PEG-maleimide. This allows selective modification at the desired location, so PEGylation does not interfere with protein function and ensures product homogeneity. To date, no long-term safety concerns have arisen with this method, including with PEGylated FVIII products developed by Bayer Healthcare (BAY 94-9027), Baxter (BAX 855), and Novo Nordisk (N8-GP).

Of note, PEGylated products may vary by site of PEGylation and type of PEG used. PEG molecules also may differ in size, with smaller molecules being more rapidly cleared than larger ones. In addition, tissue penetration may vary, with smaller molecules having greater permeability. With PEG molecules > 10 kDa, pinocytotic uptake into macrophages and Kupffer cells is increased; with PEG molecules > 30 kDa, kidney clearance is decreased; and with PEG molecules > 50 kDa, liver clearance is increased.^{15,16} Therefore, the pharmacokinetics and pharmacodynamics—and safety and efficacy—of different PEG solutions vary.

In addition to improving half-life, PEGylation may reduce immunogenicity. PEGylation of L491C in the A2-domain of FVIII resulted in reduced inhibitory activity of a monoclonal antibody that reacts to this highly immunogenic region of FVIII.¹⁷ This finding has been supported by other studies with different PEGylated proteins.^{15,18} PEGylation also may limit the inhibitory effect of alloantibodies.

Fusion Proteins

By covalently fusing coagulation factors with proteins having a much longer

half-life, such as immunoglobulin G1 (IgG1) or albumin, the half-life of the resulting molecule may be extended. Furthermore, the Fc domain can improve the solubility and stability of the partner molecule, allowing easy and cost-effective purification by protein-A/G affinity chromatography.¹⁹ Support for this concept comes from the results of preclinical and phase 1 studies showing that fusion of the monomeric form of the IgG1 Fc to human FVIII, FIX, and factor VIIa (FVIIa) increases the half-life approximately 1.5- to 4-fold without impairing efficacy. No adverse events have been reported.^{20,21}

Another fusion approach covalently attaches the clotting factor to albumin. Data from both preclinical and phase 1 studies have shown a 1.5- to 3-fold increase in plasma half-life and preserved efficacy with rFIX-albumin.^{22,23} This improvement was not found with recombinant albumin-bound FVIII, however, possibly because of interactions with von Willebrand factor (vWF).

Novel FVIII Molecules

An alternative strategy uses a recombinant, single-chain FVIII protein that prevents dissociation of the two chains of FVIII, resulting in higher affinity for vWF. Because vWF protects FVIII from proteolysis and clearance, this strategy improves the stability and half-life of FVIII, as supported by animal studies; this molecule now is being tested in human trials.²⁴ Other strategies focus on reducing the immunogenicity of recombinant FVIII molecules by changing or substituting glycan and sulfhydryl groups.²⁵

Novel Products in Preclinical Development

In addition to modifications of FVIII and FIX and administration of activated FVII (FVIIa), several additional novel approaches are in preclinical development, including factor Xase mimetics and inhibition of antithrombotic pathways.

FVIII enhances FIXa-mediated cleavage of FX; factor Xase mimetics accomplish this same function independent of FVIII. For example, a humanized bispecific monoclonal antibody to FIXa and

FX showed both efficacy and a prolonged half-life (2 weeks) in a simian model of acquired hemophilia.²⁶ The ultimate goal is to engineer such a product that combines efficacy, a prolonged half-life, and ease of administration via subcutaneous injection or oral delivery.

Whereas one approach of these new agents is to enhance clotting, the desired effect, hemostasis, also may be achieved by inhibiting antithrombotic pathways. For example, tissue factor pathway inhibitor (TFPI) inhibits FVIIa and FXa and helps to foster hemostasis. This concept was demonstrated by a monoclonal antibody to one of the Kunitz domains of TFPI in a rabbit model of hemophilia that resulted in decreased bleeding.²⁷ Other TFPI inhibitors include nucleic acid aptamers,²⁸ non-anticoagulant sulfated polysaccharide,²⁹ and small antagonist peptides.³⁰ Further studies will be needed to evaluate the relative merits of these approaches.

Antithrombin 3 (AT3) is another antithrombotic target. Alnylam Pharmaceuticals has developed a synthetic GalNAc-conjugated RNA interference (ALN-AT3) that suppresses liver production of AT3 mRNA. Pharmacologic studies have shown dose-dependent and reversible reduction of circulating AT3, which was associated with significantly increased thrombin generation and enhanced homeostasis in murine models of hemophilia.¹⁹ Use in wild-type animals induced thrombotic events and disseminated intravascular coagulation, whereas similar or even higher doses were well tolerated in mice with hemophilia A and B with no evidence of thrombosis.

Summary

Considerable advances in treating hemophilia and developing clotting factors have been reported over the past two decades. Current approaches focus on improving the safety, tolerability, and delivery of both existing recombinant factors and novel alternatives that target other aspects of the coagulation/antithrombotic pathways. These products are making their way from preclinical development in animal models to clinical trials in humans, offering the promise of

even greater advances for patient care in the years to come.

■ LONG-LASTING RECOMBINANT FACTOR VIII PROTEINS FOR HEMOPHILIA A

Based on a presentation by Amy D. Shapiro, MD, Chief Executive Officer and Co-Medical Director, Indiana Hemophilia Treatment Center, Indianapolis, Indiana.

PEGylation and fusion of Ig or albumin to FVIII and FIX are exciting strategies for increasing the half-life of these coagulation factors that are now being tested in phase 1–3 clinical trials. This section will review these developments.

Clinical Studies of PEG-FVIII Conjugates

Three PEG-FVIII conjugates currently are being tested in clinical trials: B-domain deleted recombinant FVIII (PEG-BDD-rFVIII; BAY 94-9027; www.clinicaltrials.gov ID No. NCT01184820), PEGylated full-length rFVIII (BAX 855; www.clinicaltrials.gov ID No. NCT01736475), and glycol-PEGylated rFVIII (N8-GP; www.clinicaltrials.gov ID No. NCT01480180).

PEGylation can impair a protein's activity if bound to the wrong site, so it is important that this link is produced in such a way that it maintains the protein's original function. One method of ensuring site-specific PEGylation is site-specific mutagenesis, which introduces cysteine mutations on the surface of B-domain deleted FVIII. Bayer HealthCare used this method in designing BAY 94-9027: PEG was conjugated to surface-exposed cysteines of rFVIII to retain full in vitro activity and vWF binding.¹⁷

BAY 94-9027 was evaluated in a phase 1 study of 14 patients with severe hemophilia A.¹⁵ Seven patients were given 25 IU/kg twice weekly, and the other seven received 60 IU/kg once weekly. BAY 94-9027 was well tolerated and effective without causing serious adverse events or immunogenicity. The half-life was 19 hours, representing a 1.6-fold increase over the half-life of standard rFVIII (approximately 12 hours). Phase 2/3 studies are underway.

An alternative strategy was employed by Baxter scientists in the design of BAX

855, a 20-kDa PEGylated full-length rFVIII. This conjugation process resulted from the combination of an activated PEG reagent with accessible amino groups on FVIII; it was optimized to target and modify mainly the ε-amino groups of lysine residues.¹⁸ BAX 855 reportedly retains all physiologic properties of FVIII except binding to the low-density lipoprotein receptor-related protein clearance receptor; preclinical testing revealed normal activity and a prolonged half-life when compared with unmodified rFVIII.

N8-GP is a rFVIII with site-directed glycoPEGylation being developed by Novo Nordisk.³¹ It is synthesized in a Chinese hamster ovary cell line with a truncated B domain of 21 amino acids; the terminal sialic acid on an O-glycan structure in the truncated B-domain is replaced by a conjugated sialic acid containing a branched 40-kDa PEG, resulting in a protein with a single medium-weight PEG attached to the B-domain.³² When the coagulation system is activated, thrombin cleaves the B-domain with the attached PEG, resulting in activated FVIII.

N8-GP was evaluated in a dose-escalation study (25, 50, or 75 IU/kg/dose) in 26 previously treated patients with severe hemophilia A.³³ It was well tolerated at all dose levels, and no patient developed an inhibitor or binding antibodies to FVIII or N8-GP. N8-GP exhibited a dose-linear pharmacokinetic profile with a mean half-life of 19 hours (range, 11.6–27.3 hours), representing a 1.6-fold increase over that of standard rFVIII.³⁴ Clearance was reduced by 30%, and the volumes of distributions were similar to those of standard products. N8-GP currently is being tested in phase 3 clinical trials. A similar product for hemophilia B, N9-GP (glycoPEGylation of rFIX), also has shown promise in clinical trials.³⁵

Fc Fusion Proteins

rFVIII-Fc was developed by Biogen Idec by fusing a single B-domain deleted rFVIII that is produced from human embryonic kidney cells to the dimeric Fc region of IgG1 without intervening linker sequences.^{36,37} Studies of rFVIII-Fc demonstrated prolonged in vivo activity when

compared with existing rFVIII products and comparable binding to vWF, with animal studies showing an approximate two-fold increase in half-life over rFVIII in murine and canine models of hemophilia. Interestingly, the half-life of rFVIII-Fc was comparable to that of rFVIII in neonatal Fc receptor (FcRn) knockout mice, supporting the role of the Fc fragment and interaction with FcRn in protecting the fusion protein from degradation.³⁸

A phase 1/2 study of rFVIII-Fc in 16 patients with severe hemophilia who were given either 25 or 65 IU/kg rFVIII followed by an equal dose of rFVIII-Fc showed a 1.5- to 1.7-fold increase in mean half-life (18.8 hours for both doses) for rFVIII-Fc over that of rFVIII (12.2 hours for the lower dose and 11.0 hours for the

PEGylation and Fc fusion are exciting strategies that may prolong the half-life of rFVIII, extend dosing intervals, and potentially improve compliance, access, and safety.

higher dose).²⁰ Both products had similar dose-dependent peak plasma concentrations. No drug-related adverse events, inhibitors, or severe bleeding was observed.

A phase 3, multicenter study of rFVIII-Fc (A-LONG, www.ClinicalTrials.gov ID No. NCT01181128) was completed recently. Treatment arms included individualized prophylaxis at 3- to 5-day intervals, weekly prophylaxis, and episodic (on-demand) treatment. Preliminary results from this study were presented by Mahlangu and coworkers³⁹ at the 65th Annual Meeting of the National Hemophilia Foundation in October 2013 and are summarized by Dr. Holleh D. Hussein elsewhere in this edition of *The Hemophilia Report*. Once the data from

this trial are fully compiled and analyzed, the study should provide valuable information on the safety and effectiveness of different strategies for prophylaxis of hemophilia A and on-demand treatment of bleeding episodes with rFVIII-Fc. A similar study is ongoing in children (Kids A-LONG, www.ClinicalTrials.gov identifier NCT01458106).

Summary

PEGylation and Fc fusion are exciting strategies that may prolong the half-life of rFVIII; extend dosing intervals; and potentially improve compliance, access, and safety. Both PEGylated and Fc fusion products have a half-life of 18–19 hours, whereas the half-life of vWF also is 18 hours.⁴⁰ Given that vWF is needed to stabilize and protect FVIII, it is possible that the half-life of vWF may represent a new limit to how far the half-life of VIII products may be extended.^{11,17,38} This remains a significant improvement over conventional products, and the final results of phase 3 studies such as A-LONG are eagerly anticipated.

■ THE OLD AND THE NEW: PCCs, rFVIIa, AND LONG-LASTING COAGULATION PROTEINS

Based on a presentation by Margaret V. Ragni, MD, MPH, Professor of Medicine, Division of Hematology/Oncology, University of Pittsburgh, and Director, Hemophilia Center of Western Pennsylvania, Pittsburgh, Pennsylvania.

PCCs contain combinations of clotting factors and proteins C and S. Four-factor PCCs (eg, Beriplex, Octaplex, Kcentra) contain factors II, VII, IX, and X, whereas three-factor PCCs (eg, Bebulin, Proflinone) contain factors II, IX, and X but little VII. These products initially were developed as bypassing agents to treat hemophilia patients with inhibitors to FVIII or FIX. These factors act downstream of FVIII and FIX (Figure 1), bypassing their activity. FEIBA is a formulation of PCCs that has activated clotting factors to enhance hemostasis. PCCs and FEIBA start working within minutes and carry a low risk of infectious transmission due to viral inactivation by filtration, nanofiltration, pasteurization, or solvent detergent treatment.

The substance known as rFVIIa also

was developed as a bypassing agent to treat hemophilia-associated bleeding. It may produce a “thrombin burst” via activation of FIX, FX, and FII on the surface of activated platelets. Like PCCs and FEIBA, rFVIIa is expensive, and its use carries a significant risk of thrombosis.

To assess the efficacy of these hemostatic strategies, quantitative and qualitative laboratory measures of clot formation are needed. These analytic tools include the thrombin generation assay (TGA), thromboelastography (TEG), and rotational thromboelastometry (ROTEM), which provide a more comprehensive assessment of clot formation than do such standard assays as prothrombin time (PT) and activated partial thromboplastin time (aPTT).⁴¹ Important parameters of TGA include the lag time (the time to initiation of thrombin generation) and endogenous thrombin potential (area under the curve). Those of TEG include the rate of clot formation and strength and stability of the clot. For ROTEM, important parameters include the time to clot formation (clotting time), maximum clot firmness, and time to clot lysis.

PCCs and rFVIIa in Surgery and Trauma (With or Without Warfarin)

Preclinical studies in porcine liver laceration and spleen injury models provide *in vitro* and *in vivo* evidence that PCCs are more effective than rFVIIa in restoring thrombin generation and reducing blood loss.^{42,43} These results are supported by clinical studies of coagulopathic patients undergoing surgery with excessive bleeding requiring PCCs or rFVIIa.

In a study of patients undergoing cardiopulmonary bypass, three-factor PCCs (18.9–30.9 U/kg) reduced transfusion requirements to a greater degree than did rFVIIa (90–120 µg/kg).⁴⁴ Another comparison study of three-factor PCCs (25 U/kg) with rFVIIa (90 µg/kg) in 85 traumatic brain injury patients revealed significantly greater reductions in red blood cell (RBC) and FFP requirements and a lower mortality among the PCC group.⁴⁵ In a randomized trial of patients with acute major surgical hemorrhage who also received vitamin K, four-factor

PCCs were superior to FFP, whereas other studies showed that the combination of FFP and PCCs may yield even better results.^{46,47}

At the same time, in coagulopathic trauma patients (half of whom were receiving warfarin), administration of three-factor PCCs (25 U/kg) rapidly corrected the international normalized ratio (INR) and reduced the RBC requirement; however, there was no survival benefit, as with most studies of rFVIIa in trauma.⁴⁸ Controversy remains regarding rFVIIa use, given its high cost, lack of dosing guidelines, and thrombosis risk.

Another challenge of using PCCs and rFVIIa is determining the proper dose. Some studies have used algorithms that adjusted the dose based on INR (eg, three-factor PCC dosed at 25 U/kg for an INR of 2.0–3.9, 35 U/kg for an INR of 4.0–6.0, and 50 U/kg for an INR > 6.0), whereas others have used ROTEM testing to guide PCC therapy, and still others have used fixed doses.^{45,49} The optimal timing and frequency of administration also are unclear.

Of note, four-factor PCCs appear to be more effective than are three-factor PCCs.⁴⁷ This may be due to consumption of FVII in cases of extensive surgery or trauma, bleeding, or warfarin use and the fact that three-factor PCCs are poor in FVII. When the INR > 6.0, three-factor PCCs may have little efficacy.^{50,51} Interestingly, in cases of warfarin reversal for acute bleeding, administration of 10–90 µg/kg of rFVIIa rapidly corrected the INR; unlike with four-factor PCCs, however, it seemed to do little to reduce bleeding.⁵² In parallel with these findings, ROTEM clot stability and clot lysis time in warfarin-treated patients appeared to improve more after treatment with PCCs than after use of rFVIIa.⁵³

PCCs, rFVIIa, and the New Oral Anticoagulants

New oral anticoagulants (thrombin and factor Xa inhibitors) have many advantages over warfarin, including no requirement for monitoring, few drug-drug interactions, and lower bleeding rates. However, reversal of these agents for

life-threatening bleeding is complicated by the absence of an effective antidote. The half-life and duration of action of new oral anticoagulants are short. However, acute bleeding often occurs, and waiting for the drug to wear off is unacceptable.

Unfortunately, there is little evidence of the best approach to stop bleeding in patients on new oral anticoagulants. In preclinical studies, FEIBA, PCCs, and rFVIIa improved parameters such as bleeding time, PT, and aPTT, but there was poor correlation with the amount of blood loss.^{54,55} The best results appeared to follow the use of high-dose four-factor PCCs (eg, 50 U/kg) and FEIBA, with rFVIIa and FFP having little effect.^{56,57} Studies in healthy human volunteers appeared to support these findings, with reversal of abnormal TEG and ROTEM results seen with FEIBA (20–120 U/kg) and four-factor PCCs (50 U/kg); use of rFVIIa (20–120 µg/kg) was less effective.⁵⁸ However, few data for these products exist in bleeding patients.

Novel antidotes to reverse new oral anticoagulants are being developed. They include an Xa congener that neutralizes Xa coagulation inhibitor function⁵⁹ and a dabigatran-specific antidote (aDabi-Fab) that mimics thrombin structure (but not function) and binds to dabigatran with 350-fold greater avidity.⁶⁰ These agents are still in the exploratory stage.

Treatment Considerations

To reverse warfarin-related bleeding or surgical and trauma-related bleeding, PCCs appear to be superior to rFVIIa and warfarin.⁶¹ The combination of PCC and FFP or PCC and rFVIIa may correct laboratory abnormalities such as INR even more rapidly,⁴⁶ yet extreme caution must be exercised, given the significant thrombotic risk of each agent, and recommendations regarding combination therapy must await further studies. Furthermore, use of these agents should be avoided in patients with recent (< 3 months) thromboembolism.⁶¹ Even in the absence of thromboembolism, dosing should be judicious (eg, 25 U/kg of four-factor PCC for an INR of 2.0–3.9, 35 U/kg for an INR of 4.0–6.0, and 50 U/kg for an INR > 6.0).^{50,62} The lowest effective

dose of rFVIIa has not been established.

Regarding new oral anticoagulants, it is unclear whether PCCs or rFVIIa can reverse their effects. Caution should be exercised, since there apparently is little correlation between correction of laboratory abnormalities and reversal of bleeding, and dosing and monitoring are not established.^{58,63–65} New antidotes are under development but are experimental at this time. Transfusion support and surgical hemostasis should be provided, if indicated. Of note, dialysis with activated charcoal may be effective for dabigatran if initiated within 2–4 hours of ingestion; however, this may not be effective to reverse the effects of rivaroxaban, which is highly protein bound.^{65,66}

CONCLUSION

Tremendous advances have been made in hemostasis over the past several years. Advances in clotting factors (eg, PEGylation, Ig or albumin fusion with FVIII or FIX) promise to improve hemophilia treatment by increasing factor half-life, decreasing the frequency of infusions, and potentially improving compliance and access while decreasing the risk of bleeding complications. Other novel agents (eg, TFPI, AT3 inhibitors) are also very exciting. Meanwhile, FEIBA, PCCs, and rFVIIa may help decrease life-threatening bleeding for hemophilia patients and those on warfarin or new oral anticoagulants who experience surgical or traumatic bleeding. As with all these agents, caution must be exercised, given the risk of upsetting the balance between hemostasis and thrombosis. Nonetheless, the coming years promise major advances in clotting factor treatments.

REFERENCES

- Otto JC. An account of an hemorrhagic disposition existing in certain families. *Clin Orthop Relat Res*. 1996;(328):4–6.
- Lane S. Haemorrhagic diathesis: successful transduction of blood. *Lancet*. 1840;1:185–188.
- Patek AJ, Taylor FH. Hemophilia, pt II: some properties of a substance obtained from normal human plasma effective in accelerating the coagulation of hemophilic blood. *J Clin Invest*. 1937;16:113–124.
- Kingdon HS, Lundblad RL. An adventure in biotechnology: the development of haemophilia A therapeutics—from whole-blood transfusion to recombinant DNA to gene therapy. *Biotechnol Appl Biochem*. 2002;35:141–148.
- Nilsson IM, Berntorp E, Löfqvist T, Pettersson H. Twenty-five years' experience of prophylactic treatment in severe haemophilia A and B. *J Intern Med*. 1992;232:25–32.
- Manco-Johnson MJ, Abshire TC, Shapiro AD, et al. Prophylaxis versus episodic treatment to prevent joint disease in boys with severe hemophilia. *N Engl J Med*. 2007;357:535–544.
- Mondorf W, Kalnins W, Klamroth R. Patient-reported outcomes of 182 adults with severe haemophilia in Germany comparing prophylactic vs. on-demand replacement therapy. *Haemophilia*. 2013;19:558–563.
- Astermark J. Basic aspects of inhibitors to factors VIII and IX and the influence of non-genetic risk factors. *Haemophilia*. 2006;12(Suppl 6):8–13.
- Nathwani AC, Tuddenham EGD, Rangarajan S, et al. Adenovirus-associated virus vector-mediated gene transfer in hemophilia B. *N Engl J Med*. 2011;365:2357–2365.
- Powell JS. Liposomal approach towards the development of a longer-acting factor VIII. *Haemophilia*. 2007;13:23–28.
- Powell J, Martinowitz U, Windyga J, et al. Efficacy and safety of prophylaxis with once-weekly BAY 79-4980 compared with thrice-weekly rFVIII-FS in haemophilia A patients: a randomised, active-controlled, double-blind study. LipLong Study Investigators. *Thromb Haemost*. 2012;108:913–922.
- Powell JS, Nugent DJ, Harrison JA, et al. Safety and pharmacokinetics of a recombinant factor VIII with pegylated liposomes in severe hemophilia A. *J Thromb Haemost*. 2008;6:277–283.
- Alconcel SNS, Baas AS, Maynard HD. FDA-approved poly(ethylene glycol)-protein conjugate drugs. *Polym Chem*. 2011;2:1442–1448.
- Horton S, Walsh C, Emery P. Certolizumab pegol for the treatment of rheumatoid arthritis. *Expert Opin Biol Ther*. 2012;12:235–249.
- Ivens IA, Baumann A, McDonald TA, Humphries TJ, Michaels LA, Mathew P. PEGylated therapeutic proteins for haemophilia treatment: a review for haemophilia caregivers. *Haemophilia*. 2013;19:11–20.
- Tang L, Leong L, Sim D, et al. von Willebrand factor contributes to longer half-life of PEGylated factor VIII in vivo. *Haemophilia*. 2013;19:539–545.
- Mei B, Pan J, Jiang H, et al. Rational design of a fully active, long-acting PEGylated factor VIII for hemophilia A treatment. *Blood*. 2010;116:270–279.
- Turecek PL, Bossard MJ, Graninger M, et al. BAX 855, a PEGylated rFVIII product with prolonged half-life: development, functional and structural characterisation. *Haemostaseologie*. 2012;32:S29–S38.
- Kaufman RJ, Powell JS. Molecular approaches for improved clotting factors for hemophilia. *Blood*. 2013;122:3568–3574.
- Powell JS, Josephson NC, Quon D, et al. Safety and prolonged activity of recombinant factor VIII Fc fusion protein in hemophilia A patients. *Blood*. 2012;119:3031–3037.
- Shapiro AD, Ragni MV, Valentino LA, et al. Recombinant factor IX-Fc fusion protein (rFIXFc) demonstrates safety and prolonged activity in a phase 1/2a study in hemophilia B patients. *Blood*. 2012;119:666–672.
- Weimer T, Wormsbächer W, Kronthaler U, Lang W, Liebing U, Schulte S. Prolonged in-vivo half-life of factor VIIa by fusion to albumin. *Thromb Haemost*. 2008;99:659–667.
- Santagostino P, Negrier C, Klamroth R, et al. Safety and pharmacokinetics of a novel recombinant fusion protein linking coagulation factor IX with albumin (rIX-FP) in hemophilia B patients. *Blood*. 2012;120:2405–2411.
- Schulte S. Innovative coagulation factors: albumin fusion technology and recombinant single-chain factor VIII. *Thromb Res*. 2013;131:S2–S6.
- Kannicht C, Ramström M, Kohla G, et al. Characterisation of the post-translational modifications of a novel, human cell line-derived recombinant human factor VIII. *Thromb Res*. 2013;131:78–88.
- Kitazawa T, Igawa T, Sampei Z, et al. A bispecific antibody to factors IXa and X restores factor VIII hemostatic activity in a hemophilia A model. *Nat Med*. 2012;18:1570–1574.
- Hilden I, Lauritzen B, Sørensen BB, et al. Hemostatic effect of a monoclonal antibody mAb 2021 blocking the interaction between FXa and TFPI in a rabbit hemophilia model. *Blood*. 2012;119:5871–5878.
- Gorczyca ME, Nair SC, Jilka B, et al. Inhibition of tissue factor pathway inhibitor by the aptamer BAX499 improves clotting of hemophilic blood and plasma. *J Thromb Haemost*. 2012;10:1581–1590.
- Prasad S, Lillicrap D, Labelle A, et al. Efficacy and safety of a new-class hemostatic drug candidate, AV513, in dogs with hemophilia A. *Blood*. 2008;111:672–679.
- Peraramelli S, Thomassen S, Heinzmann A, et al. Direct inhibition of factor VIIa by TFPI and TFPI constructs. *J Thromb Haemost*. 2013;11:704–714.
- Agersø H, Stennicke HR, Pelzer H, et al. Pharmacokinetics and pharmacodynamics of turoctocog alfa and N8-GP in hemophilia A dogs. *Haemophilia*. 2012;18:941–947.
- Martinowitz U, Bierre J, Brand B, et al. Bioequivalence between two serum-free recombinant factor VIII preparations (N8 and ADVATE®)—an open-label, sequential dosing pharmacokinetic study in patients with severe hemophilia A. *Haemophilia*. 2011;17:854–859.
- Tiede A, Brand B, Fischer R, et al. Enhancing the pharmacokinetic properties of recombinant factor VIII: first-in-human trial of glycoPEGylated recombinant factor VIII in patients with hemophilia A. *J Thromb Haemost*. 2013;11:670–678.
- Moss J, Rosholm A, Laurén A. Safety and pharmacokinetics of a glycoPEGylated recombinant activated factor VII derivative: a randomized first human dose trial in healthy subjects. *J Thromb Haemost*. 2011;9:1368–1374.
- Negrier C, Knoke K, Tiede A, Giangrande P, Moss J. Enhanced pharmacokinetic properties of a glycoPEGylated recombinant factor IX: a first human dose trial in patients with hemophilia B. *Blood*. 2011;118:2695–2701.

36. Peters RT, Low SC, Kamphaus GD, et al. Prolonged activity of factor IX as a monomeric Fc fusion protein. *Blood*. 2010;115:2057–2064.
37. Peters RT, Toby G, Lu Q, et al. Biochemical and functional characterization of a recombinant monomeric factor VIII-Fc fusion protein. *J Thromb Haemost*. 2013;11:132–141.
38. Dumont JA, Liu T, Low SC, et al. Prolonged activity of a recombinant factor VIII-Fc fusion protein in hemophilia A mice and dogs. *Blood*. 2012;119:3024–3030.
39. Mahlangu J, Powell J, Ragni M, et al. A-LONG: Phase 3 study of long-lasting recombinant factor VIII Fc fusion protein (rFVIII-Fc) in hemophilia A. Presented at the 65th Annual Meeting of the National Hemophilia Foundation; October 3–5, 2013; Anaheim, California. Abstract CR38.
40. Mannucci PM, Kempton C, Millar C, et al. Pharmacokinetics and safety of a novel recombinant human von Willebrand factor manufactured with a plasma-free method: a prospective clinical trial. *Blood*. 2013;122:648–657.
41. Tanaka KA, Key NS, Lew JH. Blood coagulation: hemostasis and thrombin regulation. *Anesth Analg*. 2009;108:1433–1446.
42. Mitterlechner T, Innerhofer P, Streif W, et al. Prothrombin complex concentrate and recombinant prothrombin alone or in combination with recombinant factor X and FVIIa in dilutional coagulopathy: a porcine model. *J Thromb Haemost*. 2011;9:729–737.
43. Dickneite G, Dörr B, Kasperer F, Tanaka KA. Prothrombin complex concentrate versus recombinant factor VIIa for reversal of hemodilutional coagulopathy in a porcine trauma model. *J Trauma*. 2010;68:1151–1157.
44. Tanaka KA, Mazzeffi MA, Grube M, Ogawa S, Chen EP. Three-factor prothrombin complex concentrate and hemostasis after high-risk cardiovascular surgery. *Transfusion*. 2013;53:920–921.
45. Joseph B, Jadjacharia P, Aziz H, et al. Prothrombin complex concentrate: an effective therapy in reversing the coagulopathy of traumatic brain injury. *J Trauma Acute Care Surg*. 2013;74:248–253.
46. Holland L, Warkentin TE, Refaai M, Cowther MA, Johnston MA, Sarode R. Suboptimal effect of a three-factor prothrombin complex concentrate (Profilnine-SD) in correcting supra-therapeutic international normalized ratio due to warfarin overdose. *Transfusion*. 2009;49:1171–1177.
47. Ogawa S, Szlam F, Ohnishi T, Molinaro RJ, Hosokawa K, Tanaka KA. A comparative study of prothrombin complex concentrates and fresh-frozen plasma for warfarin reversal under static and flow conditions. *Thromb Haemost*. 2011;106:1215–1223.
48. Gill R, Herbertson M, Buylsteke A, et al. Safety and efficacy of recombinant activated factor VII: a randomized placebo-controlled trial in the setting of bleeding after cardiac surgery. *Circulation*. 2009;120:21–27.
49. Weber CE, Görlinger K, Meininger D, et al. Point-of-care testing: a prospective, randomized clinical trial of efficacy in coagulopathic cardiac surgery patients. *Anesthesiology*. 2012;117:531–547.
50. Pabinger I, Brenner B, Kalina U, Knaumb S, Nagy A, Ostermann H. Prothrombin complex concentrate (Beriplex P/N) for emergency anticoagulation reversal: a prospective multinational clinical trial. Beriplex P/N Anticoagulation Reversal Study Group. *J Thromb Haemost*. 2008;6:622–631.
51. Makris M, van Veen JJ. Three or four factor prothrombin complex concentrate for emergency anticoagulation reversal? *Blood Transfus*. 2011;9:117–119.
52. Rosovsky RP, Crowther MA. What is the evidence for the off-label use of recombinant factor VIIa (rFVIIa) in the acute reversal of warfarin? *Hematology Am Soc Hematol Educ Program*. 2008:36–38.
53. Sniecinski RM, Chen EP, Makadia SS, Kikura M, Bolliger D, Tanaka KA. Changing from aprotinin to tranexamic acid results in increased use of blood products and recombinant factor VIIa for aortic surgery requiring hypothermic arrest. *J Cardiothorac Vasc Anesth*. 2010;24:959–963.
54. Imberti D, Barillari G, Biasioli C, et al. Emergency reversal of anticoagulation with a three-factor prothrombin complex concentrate in patients with intracranial haemorrhage. *Blood Transfus*. 2011;9:148–155.
55. Perzborn E, Gruber A, Tinel H, et al. Reversal of rivaroxaban anticoagulation by haemostatic agents in rats and primates. *Thromb Haemost*. 2013;110:162–172.
56. Zhou W, Schwarting S, Illanes S, et al. Hemostatic therapy in experimental intracerebral hemorrhage associated with the direct thrombin inhibitor dabigatran. *Stroke*. 2011;42:3594–3599.
57. Pragst I, Zeitler SH, Doerr B, et al. Reversal of dabigatran anticoagulation by prothrombin complex concentrate (Beriplex P/N) in a rabbit model. *J Thromb Haemost*. 2012;10:1841–1848.
58. Marlu R, Jodaj E, Paris A, Albaladejo P, Cracowski JL, Pernod G. Effect of non-specific reversal agents on anticoagulant activity of dabigatran and rivaroxaban: a randomised crossover *ex vivo* study in healthy volunteers. *Thromb Haemost*. 2012;108:217–224.
59. Lu G, DeGuzman FR, Hollenbach SJ, et al. A specific antidote for reversal of anticoagulation by direct and indirect inhibitors of coagulation factor Xa. *Nat Med*. 2013;19:446–451.
60. Schiele F, van Ryn J, Canada K, et al. A specific antidote for dabigatran: functional and structural characterization. *Blood*. 2013;121:3554–3562.
61. Ragni MV. The old and new: PCCs, VIIa, and long-lasting clotting factors for hemophilia and other bleeding disorders. *Hematology Am Soc Hematol Educ Program*. 2013:44–51.
62. Levy JH, Tanaka KA, Dietrich W. Perioperative hemostatic management of patients treated with vitamin K antagonists. *Anesthesiology*. 2008;109:918–926.
63. Eerenberg ES, Kamphuisen PW, Sijpkens MK, Meijers JC, Buller HR, Levi M. Reversal of rivaroxaban and dabigatran by prothrombin complex concentrate: a randomized, placebo-controlled, crossover study in healthy subjects. *Circulation*. 2011;124:1573–1579.
64. Weitz JI, Quinlan DJ, Eikelboom JW. Periprocedural management and approach to bleeding in patients taking dabigatran. *Circulation*. 2012;126:2428–2432.
65. Baglin T, Hillarp A, Tripodi A, Elalamy I, Buller H, Ageno W. Measuring oral direct inhibitors (ODIs) of thrombin and factor Xa: a recommendation from the Subcommittee on Control of Anticoagulation of the Scientific and Standardisation Committee of the International Society on Thrombosis and Haemostasis. *J Thromb Haemost*. 2013 Jan 24. [Epub ahead of print.]
66. Levi M, Eerenberg E, Kamphuisen PW. Bleeding risk and reversal strategies for old and new anticoagulants and antiplatelet agents. *J Thromb Haemost*. 2011;9:1705–1712.